# REVIEW

# **Apoptosis and lung injury**

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Abstract. Apoptosis is important in developmental biology and in remodeling of tissues during repair. Apoptosis also plays important roles in the progression of many diseases. The cellular and molecular mechanisms of apoptosis, in general, have been extensively demonstrated. However, the causes and the roles of apoptosis of various cell types in the lung are not well understood. We have determined that adenosine/homocysteine causes lung vascular endothelial cell apoptosis by inhibition of carboxyl methylation of the small GTPase, Ras, through inhibition of isoprenylcysteine carboxyl methyltransferase (ICMT) activity, leading to inactivation of Ras and the subsequent disruption of focal adhesion complexes, resulting in cell-extracellular matrix detachment and anoikis. Apoptosis can either ameliorate or exacerbate lung injury, depending upon the cell type. Although apoptosis of polymorphonuclear leukocytes in the lung prevents inflammation and the development of acute respiratory distress syndrome during acute lung injury, Fas/FasL-mediated alveolar epithelial cell apoptosis promotes acute lung injury and pulmonary fibrosis. Lung epithelial and endothelial cell apoptosis also contributes to the development of emphysema. This article focuses on elucidating the mechanisms of adenosine/homocysteine-induced endothelial cell apoptosis. We also review the current understanding of the role of lung cell apoptosis in acute lung injury, pulmonary fibrosis and emphysema. (Keio | Med 54 (4): 184-189, December 2005)

Key words: apoptosis, adenosine/homocysteine, acute lung injury, pulmonary fibrosis, emphysema

## **Overview of Apoptosis**

The term apoptosis, originally defined as 'falling off of leaves from trees', was first used scientifically to describe energy-dependent cell death by Kerr et al. in 1972.<sup>1</sup> Apoptosis or programmed cell death describes a genetically determined elimination of cells. The process is initiated by a death signal that tilts the balance between pro- and anti-apoptotic factors. Apoptotic cells undergo well ordered morphologic and molecular alterations, including cytoskeletal rearrangement, nuclear membrane collapse, chromatin condensation, DNA fragmentation, cell shrinkage, plasma membrane blebbing, and formation of apoptotic bodies.<sup>1</sup> In contrast, cells undergoing necrosis swell and lyse, thereby releasing intracellular contents into the interstitium leading to an inflammatory response.<sup>2</sup> Apoptosis is important in developmental biology, in remodeling of tissues during repair, and in the progression of some diseases.

#### **Cellular and Molecular Basis of Apoptosis**

Various factors, such as Fas ligand (FasL), tumor necrosis factor- $\alpha$  (TNF- $\alpha$ ), metabolite deprivation, DNA damage, or hypoxia can trigger cell apoptosis. There are two fundamental signaling pathways, the extrinsic and intrinsic pathways, by which apoptosis is mediated, as described in Fig. 1. The extrinsic pathway is activated by external death ligands. The intrinsic pathway is triggered by internal apoptotic signals and involved in mitochondria. The two pathways merge and share mechanisms utilizing the aspartate-specific cysteinyl protease (caspase) cascades.

The caspase cascade is central to the progression of apoptosis. To date, 13 mammalian caspases have been

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#### Intrinsic pathway

**Extrinsic pathway** 

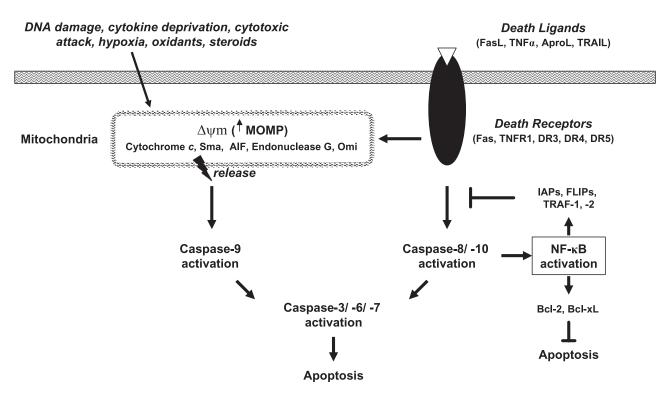


Fig. 1 Schematic representation of the cellular and molecular basis of apoptosis.

identified, which are classified into initiator caspases and effector caspases. Initiator caspases, caspase-8, -9, and -10, are activated by autoproteolysis in response to death signals and initiate apoptosis.<sup>3,4</sup> Effector caspases, caspase-3, -6, and -7, are activated by initiator caspases and cleave substrate proteins, culminating in cell death.<sup>4</sup> Over 100 caspase substrates have been identified; including inhibitor of caspase-activated DNase (ICAD), poly ADP-ribose polymerase (PARP), Bcl-2, lamin, and several cytoskeleton binding proteins. Cleavage of these proteins causes DNA fragmentation,<sup>4</sup> inhibition of DNA synthesis and repair,<sup>5</sup> nuclear membrane disruption and chromatin condensation,<sup>6</sup> and cytoskeleton collapse.<sup>7–11</sup>

### The extrinsic pathway

Binding of specific ligand to cell surface or soluble receptor initiates the extrinsic pathway and apoptosis. The best characterized ligands and their corresponding death receptors include FasL/Fas, TNF- $\alpha$ /TNF receptor 1 (TNFR1), AproL/death receptor 3 (DR3) and Apo2L (TRAIL)/death receptor 4 and 5 (DR4, DR5).<sup>12,13</sup> Upon ligand binding, adapter proteins are recruited, resulting in association and activation of initiator cas-

pases, which subsequently cleaves and activates effector caspases or activates Bid-dependent mitochondrial/intrinsic pathway<sup>14,15</sup> and apoptosis ensues.

#### The intrinsic pathway

Caspase activity is not necessary for apoptosis.<sup>16</sup> Mitochondria also play important roles in executing apoptosis. Exposure to stresses, such as cytotoxic drugs, oxidants, radiation, and growth factor deprivation, promotes mitochondrial outer membrane permeabilization (MOMP), resulting in mitochondrial release of several apoptogenic proteins to the cytosol; including cytochrome c, Smac, apoptosis-inducing factor (AIF), endonuclease G, and a serine protease called Omi.<sup>17</sup> The released cytochrome c rapidly binds to Apaf-1, leading to activation of caspase-9, with subsequent activation of caspase-3, -6, or -7, and culminating in apoptosis.18,19 Released Smac and Omi activate effector caspases by removal of inhibitor of apoptosis proteins (IAPs).<sup>20,21</sup> AIF translocates to the nucleus and initiates chromatin condensation,<sup>22</sup> and endonuclease G contributes to DNA fragmentation by cleavage of genomic DNA.23

#### Regulation of apoptosis

Development and tissue homeostasis require a proper balance between survival and apoptotic signals. Anti-apoptotic Bcl-2 family members, such as Bcl-2, Bcl-x<sub>I</sub> and Mcl-1 heterodimerize nonspecifically with pro-apoptotic Bcl-2 family members, such as Bax, Bak, Bim, and Bid, and sequester them in an inactive state in cytoplasm. This sequestration protects against apoptosis by preventing MOMP. Fas-associated death domain (FADD)-like inhibitory proteins (FLIPs) block apoptosis by competing with procaspase-8 for binding with adapter protein FADD.<sup>24</sup> In endothelial cells, downregulation of c-FLIP is implicated in extracellular matrix detachment-induced anoikis.<sup>25</sup> IAPs prevent apoptosis by binding to effector caspases, rendering them inactive.<sup>26</sup> Death signaling-induced NF-kB activation promotes expression of various anti-apoptotic proteins, including Bcl-2, Bcl-xL, FLIPs, IAP-1 and 2, TNFRassociated factor 1 and 2 to protect against apoptosis.<sup>27-29</sup>

#### **Endothelial Cell Apoptosis**

Endothelial cells form a monolayer lining the vasculature. Due to the positioning of the endothelium at the interface between the blood and surrounding tissue, the endothelium is exposed to multiple biochemical and biomechanical stresses, such as lipopolysaccharide (LPS), endotoxin, TNF- $\alpha$ , and oxidative stresses. One pathological consequence of stresses in the blood vessel is the induction of endothelial cell apoptosis. In this article, we review endothelial cell apoptosis caused by adenosine/homocysteine.

Increased levels of adenosine triphosphate (ATP) or adenosine may occur in blood vessels upon exocytotic release of nucleotides from stimulated platelet granules, during cytolytic release from necrotic cells, or from endothelial cell membrane transporters.<sup>30</sup> We have demonstrated that elevated levels of ATP or adenosine promote endothelial cell apoptosis in vitro.31,32 Ectonucleotidase-mediated hydrolysis of ATP and the subsequent uptake of adenosine were necessary for the induction of endothelial cell apoptosis.<sup>31</sup> We have also shown that adenosine-induced endothelial cell apoptosis was exacerbated by homocysteine and mimicked by inhibitors of S-adenosyl-L-homocysteine hydrolase.<sup>32</sup> In addition, adenosine- and adenosine/homocysteineinduced apoptosis correlated with protein tyrosine phosphatase-dependent inhibition of p38 mitogenactivated protein kinase (p38 MAPK)<sup>33</sup> and degradation of focal adhesion kinase (FAK), paxillin, and p130<sup>CAS</sup> proteins, with subsequent disruption of focal adhesion complexes.<sup>8</sup> Importantly, overexpression of wild type FAK blunted adenosine/homocysteineinduced endothelial cell apoptosis, while overexpression of mutants of FAK, neither FAT nor FRNK, which lack the central catalytic domain, didn't protect against adenosine- or adenosine/homocysteine-induced apoptosis.<sup>34</sup> Mutation of the autophosphorylation site of FAK also failed to protect against apoptosis,<sup>34</sup> suggesting that kinase activity is required for FAK protection.

The small GTPases are a superfamily of monomeric regulatory GTP-binding proteins. The Ras protein was the first characterized and now is known to consist of 5 major classes - Ras, Rho, Rab, Arf, and RAN. Ras GTPases are molecular switches, in that they are activated when bound to GTP, and inactivated when bound to GDP. This process is regulated by guanine nucleotide exchange factors (GEFs), GTPase-activating proteins (GAPs) and guanine nucleotide dissociation inhibitors (GDIs). GEFs accelerate the exchange rate of bound GDP for GTP. GAPs enhance hydrolysis of bound GTP to GDP, whereas GDIs inhibit the exchange of bound GDP for GTP, inhibit GTP hydrolysis, and prevent membrane association.<sup>35</sup> Our studies have demonstrated that increased adenosine/homocysteine decreased Ras GTPase activity by inhibiting carboxyl methylation of this small GTPase through inhibition isoprenylcysteine carboxyl methyltransferase of (ICMT) activity.<sup>36</sup> Transient overexpression of wild type or dominant active H-Ras blunted adenosine/ homocysteine-induced endothelial cell apoptosis,<sup>36</sup> suggesting that Ras GTPase methylation and subsequent activation play an important role in adenosine/ homocysteine-induced endothelial apoptosis.

Thus, it is speculated that increased levels of adenosine and homocysteine promote endothelial cell apoptosis by multiple signaling pathways, resulting in disruption of cell-extracellular matrix interactions and anoikis, as described in Fig. 2.

#### **Apoptosis and Lung Diseases**

The lung is a complex organ which includes many different types of cells; including endothelial cells, epithelial cells, fibroblasts, and inflammatory cells. Apoptosis can either ameliorate or exacerbate lung injury, depending upon the cell type. This review article focuses on the effects of apoptosis on acute lung injury, pulmonary fibrosis, and emphysema.

#### Apoptosis and acute lung injury

#### PMN may infiltrate lung tissue in acute lung injury

Polymorphonuclear leukocytes (PMN) undergo apoptosis *in vitro*. Macrophages phagocytose apoptotic PMN. It has been suggested that enhanced PMN apop-

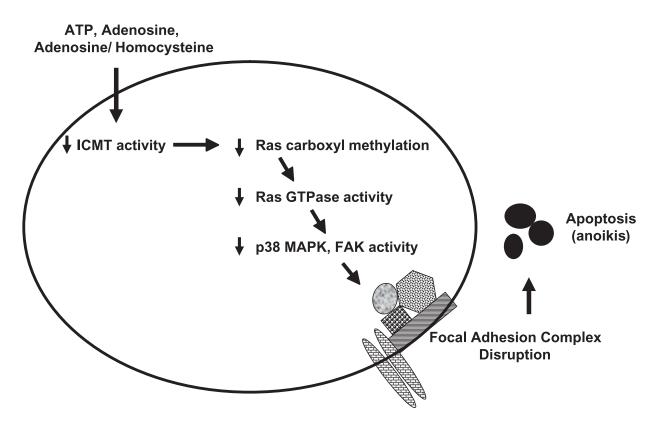


Fig. 2 Schematic representation of the signal mechanism of adenosine/homocysteine-induced endothelial apoptosis.

tosis may blunt inflammation.<sup>37</sup> The work of Matute-Bello and colleagues has extended this observation to acute lung injury. They found that there are fewer apoptotic PMN in bronchoalveolar lavage (BAL) of patients with acute respiratory distress syndrome (ARDS) or at risk of ARDS. Furthermore, BAL from ARDS patients decreases apoptosis and prolongs survival *in vitro* of normal human PMN, due to the presence of anti-apoptotic factors, such as GM-CSF.<sup>38</sup> These observations suggest that lack of PMN apoptosis may prolong inflammatory responses and predispose the patients to ARDS after acute lung injury.

#### Fas/FasL-mediated apoptosis causes acute lung injury

The Fas/FasL pathway is an important apoptosissignaling system. Apoptosis mediated by Fas/FasL interaction has been implicated in acute lung injury (ALI) and ARDS. Fas is a 45-kDa type I membrane protein, while FasL is a 37-kDa type II protein. In the lung, Fas has been found on the surface of alveolar and bronchial epithelial cells, Clara cells, alveolar macrophages, and myofibroblasts, while FasL is expressed in neutrophils and lymphocytes. Alveolar epithelial injury in humans with ALI or ARDS is associated with local upregulation of the Fas/FasL system and activation of the apoptotic cascade in alveolar epithelial cells.<sup>39</sup> Recent studies show that Fas or FasL deficient mice have lesser degrees of acute lung injury compared with wild type mice when challenged with intrapulmonary deposition of IgG immune complexes.<sup>40</sup> Inhibition of caspase activity blunts PMN-induced acute lung injury in wild type mice.<sup>40</sup> These results suggest that Fas/FasL-induced apoptosis contributes to acute lung injury.

Fas ligand is cleaved by metalloproteinases to form a soluble form, referred to as sFasL. sFasL accumulates at sites of tissue inflammation and can initiate apoptosis of leukocytes, epithelial cells, and other lung cells. sFasL is increased in BAL and pulmonary tissue of patients with multisystem organ failure (MSOF)<sup>39</sup> and in BAL of patients with ARDS.<sup>41</sup> Importantly, BAL from patients with ARDS caused apoptosis of cultured lung epithelial cells, and this effect was inhibited by blocking the Fas/FasL system,<sup>41</sup> suggesting that sFasL can be released as a biologically active, death-inducing mediator capable of inducing epithelial apoptosis by interaction with Fas during acute lung injury. Mechanical ventilation with high tidal volumes of rabbits with ARDS causes renal and intestinal epithelial cell apoptosis and renal dysfunction, which correlated with increased plasma sFasL.42 These studies suggest that sFasL also contributes to failure of other organs in MSOF.

#### Apoptosis and pulmonary fibrosis

Pulmonary fibrosis begins with alveolitis, which progresses to excess collagen deposition and destruction of normal lung architecture. Epithelial apoptosis and necrosis are increased in lungs of patients with idiopathic pulmonary fibrosis (IPF).<sup>43</sup> Apoptosis has been thought to be a non-inflammatory means of removing injurious cells thus facilitating lung repair. However, there is increasing evidence that Fas/FasL-mediated lung epithelial apoptosis induces release of proinflammatory cytokines (such as TNF- $\alpha$  and transforming growth factor- $\beta$ 1), leading to inflammation and progression from ARDS to fibrosis.44 FasL was upregulated on inflammatory cells in BAL from patients with IPF.<sup>45</sup> Fas expression was also increased on alveolar and bronchiolar epithelial cells from patients with IPF.45 sFasL was significantly enhanced in both serum and BAL from IPF patients.<sup>46</sup> Thus, we speculate that Fas/FasLinduced epithelial cell apoptosis may stimulate lung fibrosis.

#### Apoptosis and emphysema

Emphysema is characterized by loss of alveolar capillary septal tissue, resulting in impairment of gas exchange. The imbalance of proteases and anti-proteases caused by cigarette smoke is important in the pathogenesis of emphysema.<sup>47</sup> However, recent work suggests that apoptosis may also be important in the pathogenesis of emphysema. Emphysema lungs displayed apoptosis of both epithelial and endothelial cells and decreased expression of lung vascular endothelial growth factor (VEGF) and its receptor 2 (VEGF R2).<sup>48</sup> Overexpression of active caspase-3 by intratracheal instillation causes alveolar epithelial cell apoptosis and emphysema in rats.<sup>49</sup> These studies provide direct evidence that epithelial and/or endothelial apoptosis is important in the pathogenesis of emphysema. VEGF is abundantly expressed in normal lung and promotes endothelial cell proliferation. VEGF level is decreased in emphysema,<sup>48</sup> suggesting that lack of VEGF may result in apoptosis. Indeed, blockade of VEGF receptor causes apoptosis of alveolar epithelial and endothelial cells and emphysema in vitro and in vivo.<sup>50-52</sup> These studies have expanded the clinical concept of emphysema as a disease of protease-antiprotease imbalance to include the idea that programmed cell death could also be playing a role.

#### Summary

There are a variety of stimuli that may cause apoptosis via either the extrinsic or intrinsic pathways. Epithelial and/or endothelial apoptosis contributes to acute lung injury, pulmonary fibrosis, and emphysema. However, the mechanisms of endothelial apoptosis in human disease are not well understood and further study is needed. Inhibition of apoptosis in a cell-specific and vascular bed-specific manner may be potentially therapeutic for some diseases.

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#### References

- Kerr JF, Wyllie AH, Currie AR: Apoptosis: a basic biological phenomenon with wide-ranging implications in tissue kinetics. Br J Cancer 1972; 26: 239–257
- Wyllie AH, Kerr JF, Currie AR: Cell death: the significance of apoptosis. Int Rev Cytol 1980; 68: 251–306
- Chen M, Wang J: Initiator caspases in apoptosis signaling pathways. Apoptosis 2002; 7: 313–319
- Thornberry NA, Lazebnik Y: Caspases: enemies within. Science 1998; 281: 1312–1316
- Tewari M, Quan LT, O'Rourke K, Desnoyers S, Zeng Z, Beidler DR, Poirier GG, Salvesen GS, Dixit VM: Yama/CPP32 beta, a mammalian homolog of CED-3, is a CrmA-inhibitable protease that cleaves the death substrate poly(ADP-ribose) polymerase. Cell 1995; 81: 801–809
- Taimen P, Kallajoki M: NuMA and nuclear lamins behave differently in Fas-mediated apoptosis. J Cell Sci 2003; 116: 571–583
- Cheng AC, Huang TC, Lai CS, Pan MH: Induction of apoptosis by luteolin through cleavage of Bcl-2 family in human leukemia HL-60 cells. Eur J Pharmacol 2005; 509: 1–10
- Harrington EO, Smeglin A, Newton J, Ballard G, Rounds S: Protein tyrosine phosphatase-dependent proteolysis of focal adhesion complexes in endothelial cell apoptosis. Am J Physiol Lung Cell Mol Physiol 2001; 280: L342–353
- Kook S, Shim SR, Choi SJ, Ahnn J, Kim JI, Eom SH, Jung YK, Paik SG, Song WK: Caspase-mediated cleavage of p130cas in etoposide-induced apoptotic Rat-1 cells. Mol Biol Cell 2000; 11: 929–939
- Kothakota S, Azuma T, Reinhard C, Klippel A, Tang J, Chu K, McGarry TJ, Kirschner MW, Koths K, Kwiatkowski DJ, *et al*: Caspase-3-generated fragment of gelsolin: effector of morphological change in apoptosis. Science 1997; 278: 294–298
- Rudel T, Bokoch GM: Membrane and morphological changes in apoptotic cells regulated by caspase-mediated activation of PAK2. Science 1997; 276: 1571–1574
- Ashkenazi A, Dixit VM: Death receptors: signaling and modulation. Science 1998; 281: 1305–1308
- 13. Wajant H: Death receptors. Essays Biochem 2003; 39: 53-71
- Boldin MP, Goncharov TM, Goltsev YV, Wallach D: Involvement of MACH, a novel MORT1/FADD-interacting protease, in Fas/APO-1- and TNF receptor-induced cell death. Cell 1996; 85: 803–815
- Li H, Zhu H, Xu CJ, Yuan J: Cleavage of BID by caspase 8 mediates the mitochondrial damage in the Fas pathway of apoptosis. Cell 1998; 94: 491–501
- McCarthy NJ, Whyte MK, Gilbert CS, Evan GI: Inhibition of Ced-3/ICE-related proteases does not prevent cell death induced by oncogenes, DNA damage, or the Bcl-2 homologue Bak. J Cell Biol 1997; 136: 215–227

- Hung RW, Chow AW: Dissecting the "end game": clinical relevance, molecular mechanisms and laboratory assessment of apoptosis. Clin Invest Med 2004; 27: 324–344
- Slee EA, Harte MT, Kluck RM, Wolf BB, Casiano CA, Newmeyer DD, Wang HG, Reed JC, Nicholson DW, Alnemri ES, *et al*: Ordering the cytochrome c-initiated caspase cascade: hierarchical activation of caspases-2, -3, -6, -7, -8, and -10 in a caspase-9-dependent manner. J Cell Biol 1999; 144: 281–292
- Zou H, Li Y, Liu X, Wang X: An APAF-1.cytochrome c multimeric complex is a functional apoptosome that activates procaspase-9. J Biol Chem 1999; 274: 11549–11556
- Du C, Fang M, Li Y, Li L, Wang X: Smac, a mitochondrial protein that promotes cytochrome c-dependent caspase activation by eliminating IAP inhibition. Cell 2000; 102: 33–42
- Yang QH, Church-Hajduk R, Ren J, Newton ML, Du C: Omi/ HtrA2 catalytic cleavage of inhibitor of apoptosis (IAP) irreversibly inactivates IAPs and facilitates caspase activity in apoptosis. Genes Dev 2003; 17: 1487–1496
- Susin SA, Lorenzo HK, Zamzami N, Marzo I, Snow BE, Brothers GM, Mangion J, Jacotot E, Costantini P, Loeffler M, *et al*: Molecular characterization of mitochondrial apoptosis-inducing factor. Nature 1999; 397: 441–446
- Li LY, Luo X, Wang X: Endonuclease G is an apoptotic DNase when released from mitochondria. Nature 2001; 412: 95–99
- Thome M, Schneider P, Hofmann K, Fickenscher H, Meinl E, Neipel F, Mattmann C, Burns K, Bodmer JL, Schroter M, *et al*: Viral FLICE-inhibitory proteins (FLIPs) prevent apoptosis induced by death receptors. Nature 1997; 386: 517–521
- Aoudjit F, Vuori K: Matrix attachment regulates Fas-induced apoptosis in endothelial cells: a role for c-flip and implications for anoikis. J Cell Biol 2001; 152: 633–643
- Verhagen AM, Coulson EJ, Vaux DL: Inhibitor of apoptosis proteins and their relatives: IAPs and other BIRPs. Genome Biol 2001; 2: REVIEWS3009
- Tamatani M, Che YH, Matsuzaki H, Ogawa S, Okado H, Miyake S, Mizuno T, Tohyama M: Tumor necrosis factor induces Bcl-2 and Bcl-x expression through NFkappaB activation in primary hippocampal neurons. J Biol Chem 1999; 274: 8531–8538
- Wang CY, Mayo MW, Korneluk RG, Goeddel DV, Baldwin AS Jr: NF-kappaB antiapoptosis: induction of TRAF1 and TRAF2 and c-IAP1 and c-IAP2 to suppress caspase-8 activation. Science 1998; 281: 1680–1683
- Xiao CW, Asselin E, Tsang BK: Nuclear factor kappaBmediated induction of Flice-like inhibitory protein prevents tumor necrosis factor alpha-induced apoptosis in rat granulosa cells. Biol Reprod 2002; 67: 436–441
- Dubyak GR, el-Moatassim C: Signal transduction via P2purinergic receptors for extracellular ATP and other nucleotides. Am J Physiol 1993; 265: C577–606
- Dawicki DD, Chatterjee D, Wyche J, Rounds S: Extracellular ATP and adenosine cause apoptosis of pulmonary artery endothelial cells. Am J Physiol 1997; 273: L485–494
- Rounds S, Yee WL, Dawicki DD, Harrington E, Parks N, Cutaia MV: Mechanism of extracellular ATP- and adenosine-induced apoptosis of cultured pulmonary artery endothelial cells. Am J Physiol 1998; 275: L379–388
- 33. Harrington EO, Smeglin A, Parks N, Newton J, Rounds S: Adenosine induces endothelial apoptosis by activating protein tyrosine phosphatase: a possible role of p38alpha. Am J Physiol Lung Cell Mol Physiol 2000; 279: L733–742
- Bellas RE, Harrington EO, Sheahan KL, Newton J, Marcus C, Rounds S: FAK blunts adenosine-homocysteine-induced endothelial cell apoptosis: requirement for PI 3-kinase. Am J Physiol Lung Cell Mol Physiol 2002; 282: L1135–1142
- Takai Y, Sasaki T, Matozaki T: Small GTP-binding proteins. Physiol Rev 2001; 81: 153–208

- Kramer K, Harrington EO, Lu Q, Bellas R, Newton J, Sheahan KL, Rounds S: Isoprenylcysteine carboxyl methyltransferase activity modulates endothelial cell apoptosis. Mol Biol Cell 2003; 14: 848–857
- Savill J, Hogg N, Ren Y, Haslett C: Thrombospondin cooperates with CD36 and the vitronectin receptor in macrophage recognition of neutrophils undergoing apoptosis. J Clin Invest 1992; 90: 1513–1522
- Matute-Bello G, Liles WC, Radella F 2nd, Steinberg KP, Ruzinski JT, Jonas M, Chi EY, Hudson LD, Martin TR: Neutrophil apoptosis in the acute respiratory distress syndrome. Am J Respir Crit Care Med 1997; 156: 1969–1977
- 39. Albertine KH, Soulier MF, Wang Z, Ishizaka A, Hashimoto S, Zimmerman GA, Matthay MA, Ware LB: Fas and fas ligand are up-regulated in pulmonary edema fluid and lung tissue of patients with acute lung injury and the acute respiratory distress syndrome. Am J Pathol 2002; 161: 1783–1796
- Neff TA, Guo RF, Neff SB, Sarma JV, Speyer CL, Gao H, Bernacki KD, Huber-Lang M, McGuire S, Hoesel LM, *et al*: Relationship of acute lung inflammatory injury to Fas/FasL system. Am J Pathol 2005; 166: 685–694
- Matute-Bello G, Liles WC, Steinberg KP, Kiener PA, Mongovin S, Chi EY, Jonas M, Martin TR: Soluble Fas ligand induces epithelial cell apoptosis in humans with acute lung injury (ARDS). J Immunol 1999; 163: 2217–2225
- 42. Imai Y, Parodo J, Kajikawa O, de Perrot M, Fischer S, Edwards V, Cutz E, Liu M, Keshavjee S, Martin TR, *et al*: Injurious mechanical ventilation and end-organ epithelial cell apoptosis and organ dysfunction in an experimental model of acute respiratory distress syndrome. JAMA 2003; 289: 2104–2112
- Uhal BD, Joshi I, Hughes WF, Ramos C, Pardo A, Selman M: Alveolar epithelial cell death adjacent to underlying myofibroblasts in advanced fibrotic human lung. Am J Physiol 1998; 275: L1192–1199
- Chapman HA: A Fas pathway to pulmonary fibrosis. J Clin Invest 1999; 104: 1–2
- Kuwano K, Miyazaki H, Hagimoto N, Kawasaki M, Fujita M, Kunitake R, Kaneko Y, Hara N: The involvement of Fas-Fas ligand pathway in fibrosing lung diseases. Am J Respir Cell Mol Biol 1999; 20: 53–60
- 46. Kuwano K, Kawasaki M, Maeyama T, Hagimoto N, Nakamura N, Shirakawa K, Hara N: Soluble form of fas and fas ligand in BAL fluid from patients with pulmonary fibrosis and bronchiolitis obliterans organizing pneumonia. Chest 2000; 118: 451–458
- Churg A, Wright JL: Proteases and emphysema. Curr Opin Pulm Med 2005; 11: 153–159
- Kasahara Y, Tuder RM, Cool CD, Lynch DA, Flores SC, Voelkel NF: Endothelial cell death and decreased expression of vascular endothelial growth factor and vascular endothelial growth factor receptor 2 in emphysema. Am J Respir Crit Care Med 2001; 163: 737–744
- Aoshiba K, Yokohori N, Nagai A: Alveolar wall apoptosis causes lung destruction and emphysematous changes. Am J Respir Cell Mol Biol 2003; 28: 555–562
- Kasahara Y, Tuder RM, Taraseviciene-Stewart L, Le Cras TD, Abman S, Hirth PK, Waltenberger J, Voelkel NF: Inhibition of VEGF receptors causes lung cell apoptosis and emphysema. J Clin Invest 2000; 106: 1311–1319
- Tuder RM, Kasahara Y, Voelkel NF: Inhibition of vascular endothelial growth factor receptors causes emphysema in rats Chest 2000; 117: 281S
- 52. Tuder RM, Zhen L, Cho CY, Taraseviciene-Stewart L, Kasahara Y, Salvemini D, Voelkel NF, Flores SC: Oxidative stress and apoptosis interact and cause emphysema due to vascular endo-thelial growth factor receptor blockade. Am J Respir Cell Mol Biol 2003; 29: 88–97