

# REVIEW

## Regulation of Cytokine Signaling by the SOCS and Spred Family Proteins

Akihiko Yoshimura

*Department of Microbiology and Immunology, School of Medicine, Keio University, Tokyo, Japan;  
Japan Science and Technology Agency (JST), CREST, Tokyo, Japan*

(Received for publication on September 3, 2008)

(Revised for publication on October 4, 2008)

(Accepted for publication on October 9, 2008)

### Abstract

Various cytokines are involved in the regulation of the immune system and of hematopoiesis. Most cytokines utilize the so-called JAK-STAT pathway, but others activate the Ras-ERK pathway, which is more important than the STAT pathway for the proliferation of hematopoietic cells. Dysregulation of cytokine signaling can cause a variety of diseases, including allergy, inflammation, and cancer. We have identified two important regulator families involved in cytokine signaling: the SOCS proteins and the Spred proteins. Suppressors of cytokine signaling (SOCS) proteins bind to JAK and to certain receptors, thereby suppressing further signaling events. Spred family proteins interact with Ras and Raf, thereby suppressing ERK activation. Studies have shown that SOCS and Spred proteins are key physiological regulators of immunity, hematopoiesis, and angiogenesis. Evidence is also emerging for the involvement of these proteins in human diseases. (Keio J Med 58 (2) : 73–83, June 2009)

**Keywords:** cytokine, signal transduction, negative regulation

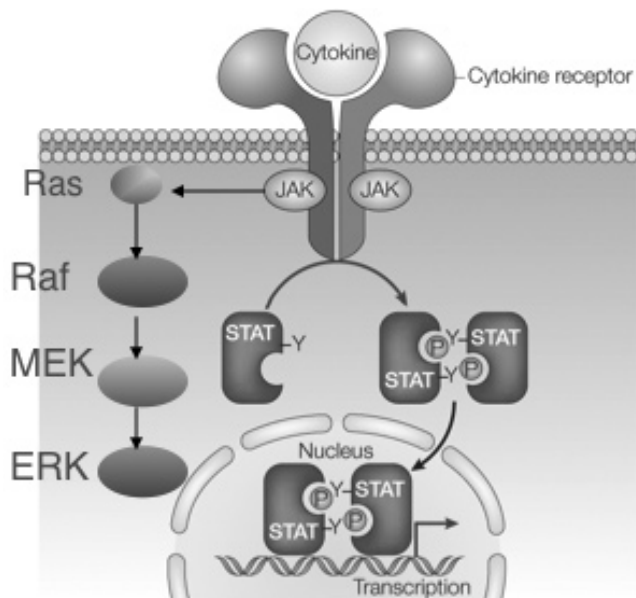
### Introduction

Cytokines play several essential roles in the development, differentiation, and function of myeloid and lymphoid cells. Some of them, including interleukins, interferons (IFNs), and hematopoietic growth factors, activate the Janus kinase/Signal Transducers and Activators of Transcription (JAK/STAT) pathway. In this pathway, cytokine binding induces receptor oligomerization, which initiates signaling from cytokine receptors. This signaling brings associated JAK kinases (JAK1, JAK2, JAK3, and Tyk2) into close apposition and allows their cross-phosphorylation and activation (**Fig. 1**).<sup>1,2</sup> The activated JAKs phosphorylate the receptor cytoplasmic domains, which creates docking sites for SH2-containing signaling proteins. Among the substrates of tyrosine phosphorylation are members of the Signal Transducers and Activators of Transcription family of proteins (STATs).<sup>1,2</sup> Although this pathway was initially found to be activated by IFNs, it is now known that a large number of cytokines, growth factors, and hormonal factors also activate

JAK and/or STAT proteins; for example, pro-inflammatory cytokine IL-6 binds to the IL-6 receptor  $\alpha$  chain and to gp130, both of which mainly activate JAK1 and STAT3. IFN $\gamma$  utilizes JAK1 and JAK2, although it mainly activates STAT1. Interestingly, anti-inflammatory cytokine IL-10 also activates STAT3. STAT4 and STAT6 are essential for Th1 and Th2 development since these are activated by IL-12 and IL-4, respectively.

In addition, the Ras-ERK pathway is activated through adaptor proteins such as SHP2 and Gab-1 (**Fig. 1**). This pathway is essential for the proliferation of hematopoietic cells through hematopoietins including IL-3, IL-5, erythropoietin (EPO), and granulocyte colony-stimulating factor (G-CSF).<sup>3,4</sup> For example, STAT3 activated by G-CSF suppresses granulopoiesis induced by G-CSF, while the Ras-ERK pathway is shown to play an essential role in promoting proliferation and anti-apoptosis of granulocyte progenitors.<sup>5</sup>

Although our understanding of the intracellular signaling molecules that mediate the functional outcome of cytokine-receptor activation has increased profoundly, the



**Fig. 1 Signal transduction of the cytokine receptors**

Most cytokines activate receptor-associated JAK tyrosine kinases, thereby activating STAT transcription factors and the Ras-ERK pathway.

most recent research has placed increasing emphasis on the mechanisms by which cytokine signals are terminated. A number of mechanisms have been proposed, including tyrosine phosphatases and transcription suppressors. In addition to these, we have previously identified two more potential mechanisms: the large CIS/SOCS and Spred/Sprouty families of proteins.<sup>6,7</sup> At the time of their discovery, the SOCS proteins were recognized as an important mechanism for the negative regulation of the cytokine-JAK-STAT pathway, but recent studies using gene-disrupted (KO) mice have revealed that they play additional, unexpected, and profound roles in many immunological processes. The Spred/Sprouty family proteins, meanwhile, are more specific to the Ras-ERK pathway, although it has been shown that they are very important regulators for development, angiogenesis and neural networks. Furthermore, their relationship to human diseases has been recently discovered.

### CIS/SOCS Family

Suppressor of cytokine signaling (SOCS) proteins and cytokine inducible SH2-containing (CIS; also known as CISH) protein molecules comprise a family of intracellular proteins, several of which have been shown to regulate the responses of immune cells to cytokines.<sup>8-10</sup> There are eight CIS/SOCS family proteins; CIS, SOCS1, SOCS2, SOCS3, SOCS4, SOCS5, SOCS6, and SOCS7, each of which has a central SH2 domain, an amino-ter-

минаl domain of variable length and sequence, and a carboxy-terminal 40-amino-acid module known as the SOCS box (**Fig. 2A**). The SOCS box is also found in other miscellaneous proteins. The SOCS box interacts with elongin B and elongin C, cullin 5, and the RING-finger-domain-only protein RBX2 (which recruits E2 ubiquitin-transferase).<sup>11</sup> CIS/SOCS family proteins, as well as other SOCS-box-containing molecules, probably function as E3 ubiquitin ligases and mediate the degradation of proteins that are associated with these family members through their N-terminal regions (**Fig. 2A**). The best characterized SOCS-family members are CIS, SOCS1, SOCS2 and SOCS3.

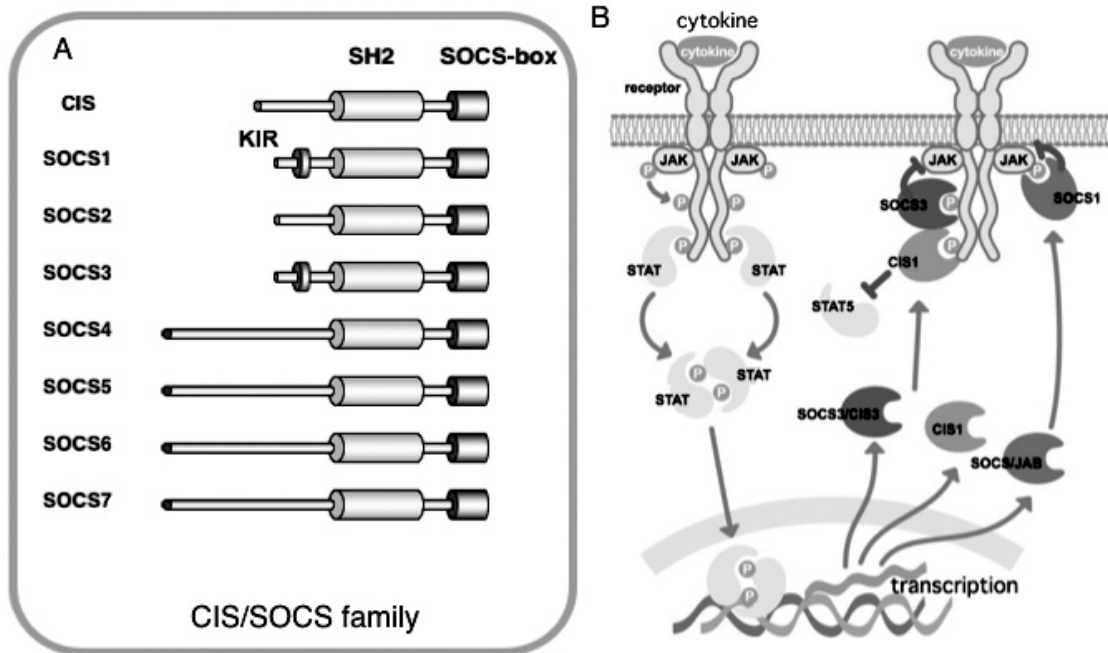
In addition to their ability to suppress signaling by ubiquitin-mediated degradation of the signaling complex, both SOCS1 and SOCS3 can inhibit JAK tyrosine kinase activity directly through their kinase inhibitory region (KIR), which has been proposed to function as a pseudosubstrate and which is important for the suppression of cytokine signals (**Fig. 2**).<sup>12</sup> The KIR peptides have been shown to inhibit JAK2-mediated phosphorylation of STAT1.

The central SH2 domain determines the target of each SOCS and CIS protein. The SH2 domains of CIS, SOCS2, and SOCS3 bind to phosphorylated tyrosine residues on activated cytokine receptors.<sup>13,14</sup> SOCS3 binds to gp130-related cytokine receptors, including the phosphorylated tyrosine 757 (Tyr757) residue of gp130 and the Tyr800 residue of IL-12 receptor  $\beta$ 2 (**Fig. 2B**).<sup>15</sup> The SH2 domain of SOCS1 directly binds to the activation loop of JAKs.<sup>12</sup> The SH2 domain of SOCS3, in contrast, does not have a high affinity to the activation loop of JAKs; yet the KIR of SOCS3 has a higher affinity to the kinase domain of JAK2 than that of SOCS1 has.<sup>16</sup> Therefore, SOCS3 might also use the same strategy of first binding with high affinity to the receptor before inhibiting JAKs through KIR (**Fig. 2B**).

### Physiological Functions of SOCS

*SOCS1 is an essential negative regulator of IFN $\gamma$*

Although *SOCS1* knockout (KO) mice are normal at birth, they exhibit stunted growth and die within 3 weeks of birth, with a syndrome characterized by severe lymphopenia, activation of peripheral T cells, fatty degeneration and necrosis of the liver, and macrophage infiltration of major organs.<sup>17</sup> The neonatal defects exhibited by *SOCS1*<sup>-/-</sup> mice appear to occur primarily as a result of unbridled IFN $\gamma$  signaling, since *SOCS1*<sup>-/-</sup> mice that also lack the IFN $\gamma$  gene or the IFN $\gamma$  receptor gene do not die neonatally.<sup>10,17</sup> Constitutive activation of STAT1 as well as constitutive expression of IFN $\gamma$ -inducible genes was observed in *SOCS1* KO mice. These data strongly suggest that the excess IFN $\gamma$  is derived from the abnormally



**Fig. 2 The structure and function of SOCS proteins**

(A) Schematic structure of the CIS/SOCS family proteins. The SOCS box is conserved in all CIS/SOCS family proteins. The function of the SOCS box is the recruitment of the ubiquitin-transferase system; thus the CIS/SOCS family proteins, as well as other SOCS-box-containing molecules, function as E3 ubiquitin ligases and mediate the degradation of proteins with which they are associated through their amino (N)-terminal regions. SOCS1 (also called JAB) and SOCS3 (also called CIS3) inhibit the tyrosine kinase activity of JAK directly, as they contain a kinase inhibitory region (KIR) immediately upstream of the central SH2 domain, which is proposed to function as a pseudosubstrate. (B) Mechanism of suppression by CIS (also called CIS1), SOCS1 (also called JAB), and SOCS3 (also called CIS3). All of these are induced by cytokine stimulation. CIS1 binds to the STAT5 activating receptors, thereby suppressing further activation of STAT5. SOCS1 binds to JAKs, and SOCS3 binds to the receptor through the SH2 domain, but both inhibit JAK activity through KIR. These complexes may be degraded by ubiquitination and proteasomal degradation recruited through the SOCS-box.

activated T cells in *SOCS1*<sup>-/-</sup> mice.

Using liver-specific *SOCS1*-conditional knockout mice, we demonstrated that *SOCS1* deletion in hepatocytes enhanced concanavalin A (ConA)-induced hepatitis, which has been shown to be dependent on NKT cells and IFN $\gamma$ .<sup>18</sup> The pro-apoptotic signals, including STAT1 and JNK activation, were enhanced in *SOCS1*-deficient mice compared to those in wild-type (WT) mice. In contrast, *SOCS1* overexpression in the liver by adenoviral gene transfer prevented ConA-induced liver injury by suppressing STAT1 activation. These findings indicate that *SOCS1* plays an important negative role in fulminant hepatitis and that forced expression of *SOCS1* is therapeutic in preventing hepatitis.<sup>18</sup>

We have recently demonstrated that *SOCS1* is essential for helper T cell differentiation. Most *SOCS1*<sup>-/-</sup>CD4 naïve T cells differentiated into Th1, even under skewing conditions, while Th17 differentiation was strongly suppressed. This was also dependent on IFN $\gamma$ , since Th17 was normally developed in *SOCS1*<sup>-/-</sup>IFN $\gamma$ <sup>-/-</sup> T cells. As a result, T cell specific *SOCS1*-deficient mice were very sensitive to dextran sulfate sodium (DSS)-induced colitis

(Th1 type disease)<sup>19</sup> but resistant to experimental autoimmune encephalomyelitis (EAE), a typical Th17 type disease.<sup>20</sup> Taken together, these data indicate that *SOCS1* negatively regulates IFN $\gamma$  signaling in various types of cells, thus fulfilling an important function for the suppression of Th1-type inflammatory diseases.

#### *Physiological functions of SOCS3 defined by gene targeting at various tissues*

*SOCS3* knockout mice die during the embryonic stage of development due to placental function defects. In other words, deletion of *SOCS3* causes embryonic lethality; these embryos can be saved, however, by a tetraploid rescue approach. These observations demonstrate *SOCS3*'s essential role in placental development and non-essential role in embryo development. Rescued *SOCS3*-deficient mouse embryos exhibit prenatal lethality with cardiac hypertrophy, which suggests that *SOCS3* is essential for regulating either LIF receptors or gp130 signaling.<sup>21</sup>

Conditional-KO mice studies have demonstrated that

*SOCS3* is an important negative regulator of IL-6<sup>15</sup> and G-CSF.<sup>22</sup> Mice in which the *SOCS3* gene was deleted in all hematopoietic cells developed neutrophilia and a spectrum of inflammatory pathologies. When stimulated with G-CSF *in vitro*, *SOCS3*-deficient cells of the neutrophilic granulocyte lineage exhibited prolonged *STAT3* activation and enhanced cellular responses to G-CSF;<sup>22</sup> *SOCS3*-deficient mice injected with G-CSF *in vivo* displayed enhanced neutrophilia, progenitor cell mobilization, and splenomegaly, but unexpectedly also developed inflammatory neutrophil infiltration into multiple tissues and consequent hind-leg paresis. Interestingly, conditional *STAT3*-deletion in neutrophils also resulted in hyper-response to G-CSF,<sup>23</sup> suggesting that a major role of *STAT3* in neutrophils is the induction of *SOCS3*. It seems likely that the ERK pathway induced by G-CSF play a major role in the proliferation and survival of neutrophils.

The essential roles of *SOCS3* in endocrine systems have also been clarified in recent years. Administration of leptin to neural cell-specific *SOCS3* conditional KO mice greatly reduces their food intake and causes enhanced body weight loss compared to WT mice, indicating that *SOCS3* in the brain negatively regulates leptin signaling.<sup>24</sup> Moreover, the *SOCS3*-deficient mice were resistant to high fat diet-induced weight gain and hyperleptinemia, and their insulin-sensitivity was retained.<sup>24,25</sup> These data indicate that *SOCS3* is a key regulator of diet-induced leptin as well as of insulin resistance. In addition, *SOCS3* deficient adipocytes generated from *SOCS3* KO fibroblasts are significantly protected from TNF- $\alpha$ -induced insulin resistance, mainly due to reduced proteasomal degradation of IRS proteins by TNF- $\alpha$ , suggesting that *SOCS3* is an important mediator of insulin resistance *in vivo*.<sup>26</sup> Consistent with these ideas is the observation that the loss of *SOCS3* in the liver apparently improved insulin sensitivity.<sup>27</sup> Unexpectedly, however, liver-specific *SOCS3* cKO mice exhibited obesity and systemic insulin resistance with age, due to constitutive activation of *STAT3* which mimics chronic inflammation. Collectively, these results indicate that *SOCS3* can be a potential therapeutic target for human metabolic disorders such as obesity and diabetes, although long-term treatment may cause inconvenient side effects.

### SOCS and Immunity

#### *The role of SOCS1 in innate immunity and inflammatory diseases*

Toll-like receptor (TLR) signals that initiate innate immune responses to pathogens must be tightly regulated to prevent excessive inflammatory damage in the host. TLR ligands, such as LPS and CpG DNA, are potent inducers of *SOCS1* and *SOCS3*; therefore the role of

*SOCS1* and *SOCS3* in TLR responses has been extensively investigated.

*SOCS1*-deficient mice are hypersensitive to LPS, which leads to increases in tumour necrosis factor (TNF) and IL-12 production.<sup>28</sup> The hyperactivation of macrophages in *SOCS1*<sup>-/-</sup> mice might be due in part to a stronger responsiveness of *SOCS1*<sup>-/-</sup> cells to IFN $\gamma$  compared with wild-type cells. However, since *IFN $\gamma$* <sup>-/-</sup>*SOCS1*<sup>-/-</sup> mice are still sensitive to LPS-induced shock, IFN $\gamma$ -independent mechanisms probably exist. A direct effect of *SOCS1* on the TLR-NF- $\kappa$ B (nuclear factor- $\kappa$ B) pathway has been proposed.<sup>29</sup> Moreover, Kimura *et al.* indicate that LPS can activate JAK2 and *STAT5*, which are involved in IL-6 induction, and that *SOCS1* selectively inhibits this process.<sup>30</sup>

*SOCS1* also negatively regulates LPS- and IL-4-induced dendritic cell (DC) maturation. *SOCS1*-deficient DCs secrete larger amounts of IFN $\gamma$ , IL-6, IL-12, and TNF in response to LPS and CpG compared with wild-type cells.<sup>31,32</sup> *SOCS1*-deficient DCs express higher levels of MHC class II and co-stimulatory molecules.<sup>32</sup> Therefore, *SOCS1* must be deeply involved in the development, maturation, and activation of DCs.

These *SOCS1*-deficient DCs seem to be responsible for the development of systemic autoimmunity in aged *SOCS1*-deficient mice. Immunization with normal DCs can activate autoreactive T cells but rarely causes autoimmune pathology, indicating that self-tolerance is still maintained in the vaccinated hosts. However, an adoptive transfer of *SOCS1*<sup>-/-</sup> DCs to wild-type recipients resulted in the induction of autoantibodies through enhanced expression of B-cell-activating factor (BAFF) by the donor DCs.<sup>31</sup> This breaking of self-tolerance might be due to hyper-production of IL-12 by *SOCS1*-deficient DCs.<sup>32</sup> These data indicate that *SOCS1* is an essential negative regulator for T-cell activation by DCs and for the maintenance of immunological tolerance.

Depletion of the *SOCS1* protein in DCs, therefore, may enhance anti-tumour immunity. Indeed, we have shown that adoptive transfer of antigen-loaded *SOCS1*<sup>-/-</sup> BMDCs can prevent B16 melanoma growth in mice.<sup>32</sup> Similarly, Chen and colleagues reported that silencing *SOCS1* using small interfering RNA (siRNA) technology in antigen-presenting DCs strongly enhanced antigen-specific anti-tumour immunity.<sup>33</sup> A transfer of DCs treated with ovalbumin (OVA)-pulsed *SOCS1*-siRNA enhanced the proliferation and function of OVA-specific cytotoxic T cells (CTLs) compared with control DCs.

TLR signaling pathways are essential for macrophage-dependent inflammatory bowel diseases, and *SOCS1* has an important role in the development of colitis. *SOCS1*<sup>-/-</sup> *Tcr $\alpha$* <sup>-/-</sup> mice develop very severe colitis and die within two months of birth.<sup>35</sup> This very severe colitis is dependent on both IFN $\gamma$  and IL-4. In addition, NF- $\kappa$ B and MAP kinases were strongly activated in the colons of

these mice; interestingly, however, this severe colitis was completely abolished when the mice were derived in germ-free conditions. This is consistent with previous observations that eventually all colitis models in mice occur in non-germ-free conditions. These findings suggest that TLR signaling is prerequisite for pro-inflammatory cytokine production and that these two signals must be mutually activated to promote colitis. Furthermore, SOCS1 has an important role in the negative regulation of both the JAK/STAT signaling cascade and the TLR pathway during the development of inflammatory bowel diseases.

### *SOCS3 and regulation of TLR signaling*

SOCS3 has now been shown to be a key regulator for the divergent activity of IL-6 and IL-10 following TLR stimulation.<sup>15</sup> IL-6 is a pro-inflammatory cytokine that assumes a progressive role in many inflammatory diseases, while IL-10 is an immunoregulatory cytokine that has potent anti-inflammatory activity, including the suppression of gene activation through TLR signaling pathways. While it was known that STAT3 is essential for the biological actions of both IL-6 and IL-10, it was unclear how these two cytokines could have such precisely opposing functions. SOCS3 protein is strongly induced by both IL-6 and IL-10 in the presence of LPS, but IL-6 signaling is selectively inhibited due to the binding of SOCS3 to the IL-6R subunit gp130 (Tyr759), but not to the IL-10 receptor (IL-10R). Therefore, STAT3 activation is transient in response to IL-6, but is sustained for a long period in response to IL-10. Furthermore, a gp130 mutant lacking the SOCS3 binding site (Tyr759Phe), as well as any heterologous cytokine receptors that are mutated to induce sustained STAT3 activation by deleting the SOCS3 binding sites, can elicit an anti-inflammatory effect indistinguishable from that induced by IL-10. Therefore, the anti-inflammatory response is a generic cytokine signaling pathway dependent on STAT3 but not unique to the IL-10. This idea is consistent with recent studies showing that constitutively activated STAT3 (STAT3c) is sufficient for the suppression of LPS-induced TNF and IL-6 production in macrophages. We proposed that this sustained activation of STAT3 is essential for the anti-inflammatory effect, while transient activation of STAT3 promotes inflammation.

### *SOCS3 and DC-mediated T-cell differentiation*

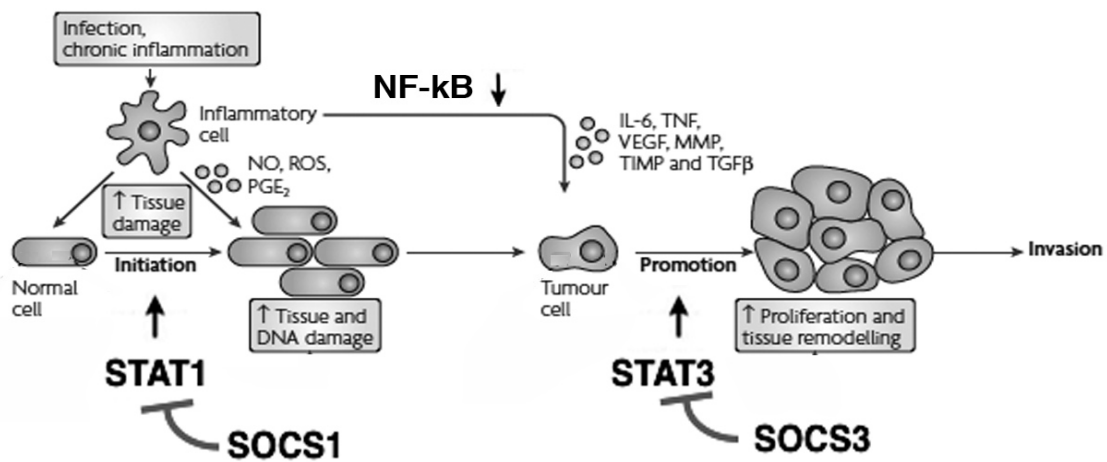
Recently, we examined the antigen-presenting cell (APC) function of *SOCS3*-deficient DCs. Like *SOCS3*<sup>-/-</sup> macrophages, *SOCS3*<sup>-/-</sup> DCs showed constitutive activation of STAT3 and expressed low levels of MHC class II molecules, co-stimulatory molecules, and IL-12.<sup>36</sup> Surprisingly, adoptive transfer of *SOCS3*<sup>-/-</sup> DCs suppressed

EAE. *SOCS3*<sup>-/-</sup> DCs are poor activators of effector CD4<sup>+</sup> T cells, but they selectively expand forkhead box P3 (FOXP3)<sup>+</sup> regulatory T cells (Tregs), which can suppress EAE. We have shown that IL-10 and TGFβ1 are target genes of STAT3.<sup>37,38</sup> FOXP3<sup>+</sup> T-cell expansion can be blocked by TGFβ-specific antibody, and indeed *SOCS3*<sup>-/-</sup> DCs produced higher levels of TGFβ than wild-type DCs did. Thus, high STAT3 activation in DCs without *SOCS3* results in increased production of IL-10 and TGFβ, thereby inducing FOXP3<sup>+</sup> Tregs. By contrast, reduced STAT3 expression due to the overexpression of SOCS3 in *SOCS3*-transduced DCs results in reduced production of IL-12, IFNγ, and IL-23, thereby inducing Th2 cells.<sup>39</sup> These results suggest that the expression of SOCS3 by DCs might have a crucial role in the balance between effector Th2 cells and Tregs.

### *SOCS3 and inflammatory diseases*

A growing body of evidence suggests that, in pathological situations, SOCS3 could suppress inflammatory reactions in which IL-6-related cytokines play important progressive roles. This is because SOCS3 is a relatively specific inhibitor of gp130, as described above. STAT3 activation and high SOCS3 expression levels have been found in epithelial and lamina propria cells in the colon of intestinal bowel disease (IBD) model mice, as well as in human ulcerative colitis and CD patients<sup>40</sup> and in synovial fibroblasts of RA patients.<sup>41</sup> STAT3 activation preceded SOCS3 expression, which is consistent with the idea that SOCS3 is part of the STAT3 negative-feedback loop.<sup>40</sup> We have shown that overexpression of SOCS3 by adenovirus gene transfer could prevent the development of experimental arthritis.<sup>41</sup> Therefore, the IL-6/STAT3 pathway promotes the progression of the chronic status of diseases by contributing to cytokine and growth factor production, tissue hyperplasia, synovial fibroblast proliferation, fibrosis, and osteoclast activation. Modulation of the gp130/JAK/STAT pathway therefore represents a reasonable strategy for new anti-inflammatory drug development. In addition, STAT3 is now known to play an essential role in the development of Th17 cells, which are extremely inflammatory and pathogenic. SOCS3 has been shown to negatively regulate Th17 development by suppressing STAT3 activated by IL-6 or IL-23.<sup>20,42</sup> Thus, again, enforced expression of SOCS3 in T cells may also ameliorate Th17-mediated inflammatory diseases such as RA.

In contrast, enhanced action of SOCS3 may promote allergic responses, since a recent analysis has indicated that transgenic SOCS3 expression in T cells inhibits Th1 development and promotes Th2 development.<sup>43</sup> Indeed, the same report also proposes that the degree to which SOCS3 expression in T cells is increased correlates with the severity of human allergic diseases such as asthma



**Fig. 3 Role of SOCS1 and SOCS3 in inflammation-associated tumourigenesis**

Two-step carcinogenesis model associated with inflammation. STAT1/SOCS1 are involved in the early stage of inflammation which leads to tissue damages, resulting in increased cellular turnover. NO and ROS from inflammatory cells may induce DNA damage, which increases the possibility of the emergence of cells possessing a high risk of malignant transformation. STAT3/SOCS3 are involved in the late stage, in which tumour promotion occurs by cellular and extracellular signals activated by cytokines from inflammatory cells or stromal cells. This step raises immortalized cells that are resistant to growth-inhibitory signals, apoptosis, and anti-tumour immunity.

and atopic dermatitis. Modulation of SOCS3 levels in T cells could help to regulate Th1/Th2 balance for the treatment of autoimmune inflammatory diseases.

### SOCS and Inflammation-associated Tumourigenesis

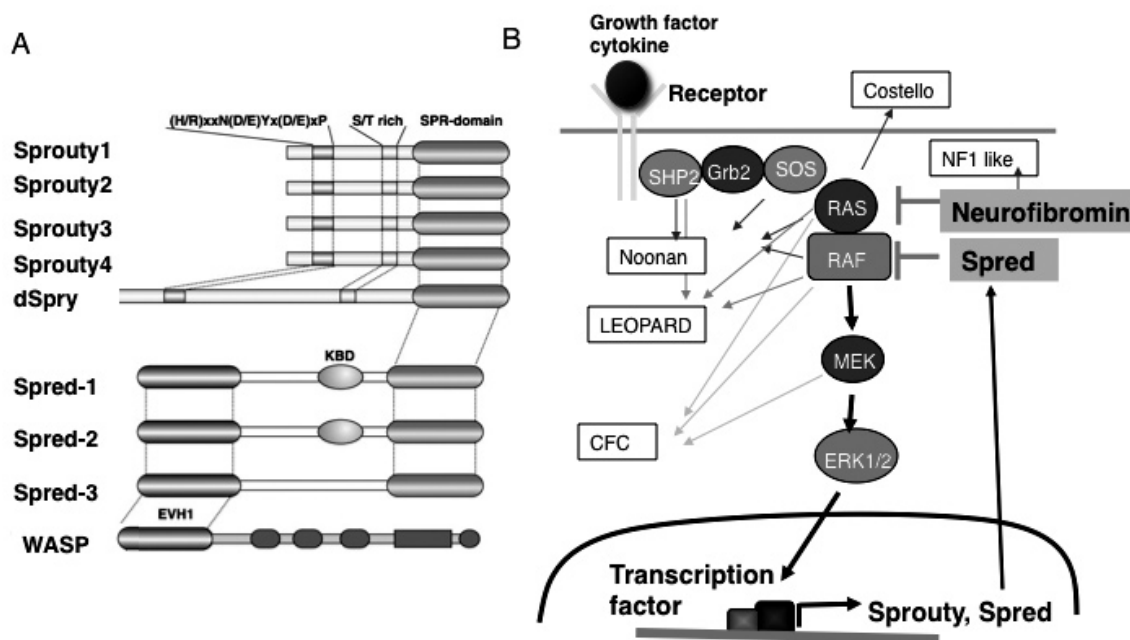
It has been estimated that more than 20% of all malignancies are initiated or exacerbated by inflammation; for example, most human hepatocellular carcinomas (HCCs) are a consequence of hepatitis C virus (HCV) infection. The expression of *SOCS1* is often silenced in these tumours by hypermethylation of CpG islands of the *SOCS1* promoter.<sup>44</sup> *SOCS1* is one of the most frequently methylated genes (65%) in HCCs, and the deletion of *SOCS1* in tumour cells might enhance IL-6-mediated cell proliferation. Therefore, *SOCS1* is considered to be an anti-oncogene because it is a suppressor of signals induced by a growth factor (IL-6). *SOCS1*<sup>+/-</sup> mice are consistently shown to be hypersensitive to dimethylnitrosamine-induced hepatocarcinogenesis.<sup>44</sup>

The full story, however, may not be so simple. We found that silencing of *SOCS1* was frequently observed even in pre-malignant HCV-infected patients.<sup>44</sup> Liver injury is associated with hyperactivation of STAT1 and reduced activation of STAT3.<sup>18,45</sup> Therefore, reduced expression of *SOCS1* might enhance tissue injury and inflammation by hyperactivation of STAT1, promoting the turnover of epithelial cells and enhancing their susceptibility to oncogenesis. The importance of *SOCS1* for in-

hibition of inflammation-associated tumour development is supported by the recent finding that *SOCS1*<sup>-/-</sup> transgenic mice, in which *SOCS1* expression is deleted in all types of cells except T and B cells, developed chronic colitis and colon tumours.<sup>46</sup> This study strongly suggests that chronic activation of the IFN $\gamma$ -STAT1 pathway that occurs in the absence of *SOCS1* causes colitis-induced colon tumours. Therefore, *SOCS1* is a unique anti-oncogene that prevents carcinogenesis by suppressing chronic inflammation (Fig. 3).

*SOCS3* might also be involved in the development and progression of malignancies. Unlike *SOCS1*, *SOCS3* expression levels were high in infected non-tumour areas of patients infected with HCV.<sup>45</sup> Reduced expression of *SOCS3* has been observed in various human cancers and is associated with constitutive STAT3 activation.<sup>47</sup> Indeed, the levels of *SOCS3* were inversely correlated with STAT3 activation in regions of human livers with and without HCC.<sup>45</sup> Numerous studies have shown that hyperactivation of STAT3 can contribute to tumourigenesis by inducing multiple tumour-promoting genes.<sup>47</sup>

Thus, we propose a two-hit model of inflammation-associated tumourigenesis (Fig. 3). In this model, initiation occurs in the form of an activating or disabling mutation in one of the molecules that regulate the cellular circuits and capacitors controlling cell division, survival, and senescence. Persistent inflammation leads to tissue damage, resulting in increased cellular turnover. Nitric oxide (NO) and reactive oxygen species (ROS) from inflammatory cells may induce DNA damage, which in-



**Fig. 4 Structure of the Spred/Sprouty family proteins and relationship to NCF1 syndrome**  
 (A) Conserved N-terminal tyrosine motifs of Sproutys and other domains are illustrated. S/T rich: serine/threonin rich region, SPR: Sprouty-related region, KBD: c-kit-binding domain, EVH1: Ena/VASP homology 1, WASP: Wiskott-Aldrich syndrome protein.  
 (B) The growth factor-induced Ras-ERK pathway is illustrated and the component proteins involved in each syndrome are shown.

creates the possibility of the emergence of cells possessing a high risk of malignant transformation. STAT1 plays a positive role in non-tumour inflammatory regions in this early stage, and *SOCS1* silencing in pre-tumour cells results in strong and persistent STAT1 activation, which induces apoptosis and tissue damage, leading to DNA damage and cell regeneration which may promote the emergence of malignant cells. Then, promotion occurs by cellular and extracellular signals activated by cytokines from inflammatory cells or stromal cells, leading to immortalized cells that are resistant to growth-inhibitory signals, apoptosis, and anti-tumour immunity. Constitutive STAT3 activation in tumour cells contributes to an expansion of tumour cells by promoting cell proliferation, survival, angiogenesis, and tissue remodeling. SOCS3 silencing is one of the mechanisms for constitutive STAT3 activation. However, the mechanism of the reduction of SOCS3 expression in tumours has not been established.

**Spred and Sprouty**

*Finding of the Spred/Sprouty family*

Sprouty was originally identified in *Drosophila* as a negative regulator of FGF (fibroblast growth factor) signaling during tracheal development. Now it is regarded

as a general inhibitor of the growth factor-induced RTK (receptor tyrosine kinase)-dependent Ras/Raf/ERK signaling pathways involved in development and organogenesis. In mammals, four Sprouty orthologues have been identified (Sprouty1, -2, -3, and -4).<sup>48</sup> In addition, we have found three Sprouty-related genes, which we call Spreds (Sprouty-related Ena/VASP homology 1 domain-containing proteins) (Fig. 4A).<sup>7,49</sup> Spreds bind to Ras and Raf, thereby suppressing activation of Raf.<sup>7</sup> Gene targeting and overexpression studies have demonstrated that mammalian Sproutys also inhibit growth factor-induced cellular responses by inhibiting the RTK-dependent ERK signaling pathway (Fig. 4B).<sup>50–52</sup>

*Spred and hematopoiesis*

Spred-1 has been implicated in hematopoiesis ever since it was observed that bone marrow-derived mast cells and eosinophils from *Spred-1*<sup>-/-</sup> mice were more sensitive to IL-3 and IL-5, respectively, than those from WT mice.<sup>53,54</sup> In *Spred-2*<sup>-/-</sup> mice, embryonic hematopoiesis in the aorta-gonad-mesonephros (AGM) region was enhanced compared with that observed in WT mice.<sup>55</sup>

*Spred-1*<sup>-/-</sup> adult animals appeared to be healthy and showed no apparent abnormalities in most organs. Studies in a murine allergic asthma model of *Spred-1*-deficient mice demonstrated that Spred-1 negatively regu-

lates allergen-induced airway eosinophilia, hyperresponsiveness, and mucus production, without affecting helper T cell differentiation. Biochemical assays demonstrated that Spred-1 suppresses IL-5-dependent cell proliferation and ERK activation. This indicates that Spred-1 negatively controls eosinophil numbers and functions by modulating IL-5 signaling in allergic asthma.<sup>54</sup>

*Spred-1* deficient mice developed myeloproliferative diseases with age. RT-PCR analysis showed high levels of Spred-1 expression in hematopoietic stem cells and bone-marrow-derived mast cells (BMMCs), erythroid cells, and B-cells, but low levels in megakaryocytes and macrophages. Spred-1 is expressed in a particular subset of mature hematopoietic cells and is inducible by IL-3. In IL-3 dependent Ba/F3 cells expressing c-kit, forced expression of Spred-1 resulted in a reduced proliferation rate and ERK activation in response to not only SCF but also IL-3.<sup>56</sup> In *Spred-1*-deficient bone-marrow-derived mast cells, proliferation and ERK/MAP kinase activation was increased in response to IL-3 or SCF.<sup>56</sup> Therefore, Spred-1 inhibits not only growth-factor-induced ERK activation but also cytokine-induced ERK activation.

#### *Spred and lymphangiogenesis*

Although the individual physiological roles of Spred-1 and Spred-2 have been investigated using gene-disrupted mice, the overlapping functions of Spred-1 and Spred-2 have not been clarified. We demonstrate that the deletion of both *Spred-1* and *Spred-2* resulted in embryonic lethality at E12.5-15.5 with marked subcutaneous hemorrhage, edema, and dilated lymphatic vessels filled with erythrocytes.<sup>57</sup> The phenotype of these mice resembled that of *Syk*<sup>-/-</sup> and *SLP-76*<sup>-/-</sup> mice, with defects in the separation of lymphatic vessels from blood vessels. The numbers of LYVE-1-positive lymphatic vessels and lymphatic endothelial cells were markedly increased in *Spred-1/2*-deficient embryos compared with wild-type embryos, while the number of blood vessels was not different. *Ex vivo* colony assay revealed that *Spred-1/2* suppressed lymphatic endothelial cell proliferation and/or differentiation. In cultured cells, the overexpression of *Spred-1* or *Spred-2* strongly suppressed vascular endothelial growth factor-C (VEGF-C)/VEGF receptor (VEGFR)-3-mediated ERK activation, while *Spred-1/2*-deficient cells were extremely sensitive to VEGFR-3 signaling. Thus, Spreds play an important role in lymphatic vessel development by negatively regulating VEGF-C/VEGFR-3 signaling. Interestingly, microRNA miR-126 has been shown to enhance the proangiogenic actions of VEGF and FGF, and to promote blood vessel formation by repressing the expression of Spred-1.<sup>58</sup> Thus, Spreds may also be involved in the regulation of angiogenesis.

#### *Spred-1 and human diseases*

Neurofibromatosis type 1 (NF1), or von Recklinghausen disease, is an autosomal dominant condition characterized by multiple café-au-lait spots, axillary freckling, Lisch nodules in the iris, and tumours of the nervous system. Other frequently observed features are short stature, macrocephaly, and learning and behavioral problems. Most NF1 is caused by inactivating mutations in the *NF1* tumour suppressor gene encoding neurofibromin, a positive regulator of RAS inactivation. NF1 was the first human disorder shown to originate from germline mutations in a gene encoding a component of the RAS-ERK pathway. Subsequently, mutations in genes encoding other components of this pathway were implicated in disorders showing some phenotypic overlap with NF1, for example, *PTPN11*, *KRAS*, and *SOS1* were implicated in Noonan syndrome, *PTPN11* in LEOPARD syndrome, *HRAS* in Costello syndrome, and *KRAS*, *BRAF*, *MEK1*, and *MEK2* in cardio-facio-cutaneous (CFC) syndrome (**Fig. 4B**). These disorders, now known as the 'neuro-cardio-facial-cutaneous' (NCFC) syndromes, present with a variable degree of cognitive impairment, facial dysmorphism, congenital heart defects, and skin abnormalities.<sup>59</sup>

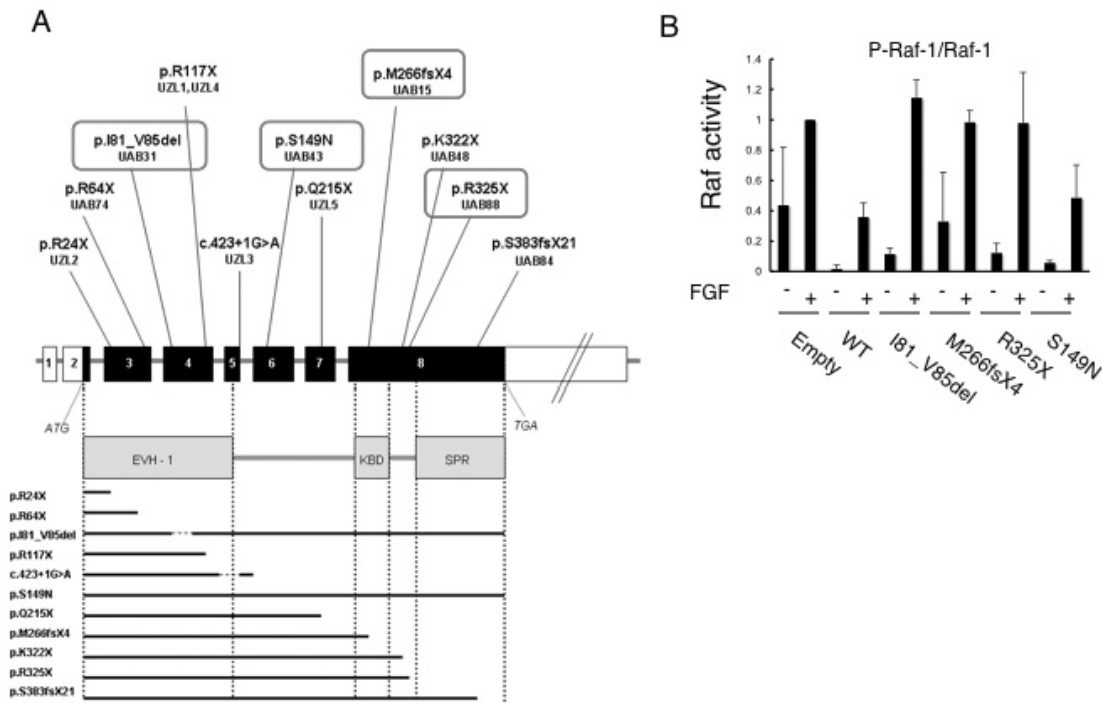
We reported germline loss-of-function mutations in *SPRED1* resulting in a newly identified autosomal dominant human disorder (**Fig. 5A**).<sup>60</sup> The clinical features of the reported disorder resemble those of neurofibromatosis type 1; they consist of multiple café-au-lait spots, axillary freckling, and macrocephaly. All mutations are found to be the loss-of-function type (**Fig. 5B**). To our knowledge, this is the first report of mutations in this family of genes causing human disease, and the existence and nature of this disorder strongly support our notion that Spred is a negative regulator of the RAS-ERK pathway.

Furthermore, Spreds are now considered to be potential tumour suppressors, as is NF-1. It has been reported that *Spred-1* and *Spred-2* expression was reduced in human HCC.<sup>61</sup> In addition, overexpression of Spred-1 can efficiently suppress tumourigenesis in nude mice.<sup>62</sup> Further study is underway to identify mutations or loss of expression of Spreds/Sproutys in cancer and to establish Spreds as a therapeutic target.

#### **Concluding Remarks**

In the past decade, following the discovery of the SOCS and Spred protein families, we have extended our understanding of the structure and function of these proteins. SOCS proteins not only act as simple negative-feedback regulators, but also play a part in the fine tuning of the immune response and in the cross-talk of the complicated cytokine signal networks. Spreds/Sproutys





**Fig. 5 Mutations in SPRED1 gene found in NF1 patients (A) and suppression of the FGF-induced Raf1 kinase activity by mutant Spred1 proteins (B).** 293 cells transfected with various mutant SPRED1 genes were stimulated with FGF, then Raf1 was immunoprecipitated. *In vitro* Raf1 kinase assay was performed using MEK1 as substrate (B).

are also very important for development, hematopoiesis, angiogenesis, and neurogenesis. Since various cytokines and stimuli are constantly in the microenvironment of immune cells, hematopoietic cells, neural cells, and endothelial cells, signal regulation by those proteins and other regulators must be important to maintain the proper homeostasis. Therefore, further investigation into the function of SOCS and Spred proteins might provide us with an unexpected role for these proteins in the regulation of signaling pathways in life.

**Acknowledgements**

We thank Ms. Y. Nishi for preparing the manuscript. This work was supported by special grants-in-aid from the Ministry of Education, Science, Technology, Sports, and Culture of Japan and the Program for Promotion of Fundamental Studies in Health Sciences of the National Institute of Biomedical Innovation (NIBIO).

**References**

1. Ward AC, Touw I, Yoshimura A: The jak-stat pathway in normal and perturbed hematopoiesis. *Blood* 2000; **95**: 19–29
2. Hanada T, Yoshimura A: Regulation of cytokine signaling and inflammation. *Cytokine and Growth Factor Reviews* 2002; **13**: 413–421

3. Hara T, Miyajima A: Function and signal transduction mediated by the interleukin 3 receptor system in hematopoiesis. *Stem Cells* 1996; **14**: 605–618
4. Yoshimura A, Misawa H: Physiology and function of the erythropoietin receptor. *Curr Opin Hematol* 1998; **5**:171–176
5. Miranda MB, Xu H, Torchia JA, Johnson DE: Cytokine-induced myeloid differentiation is dependent on activation of the MEK/ERK pathway. *Leuk Res* 2005; **29**: 1293–1306
6. Endo TA, Masuhara M, Yokouchi M, Suzuki R, Sakamoto H, Mitsui K, Matsumoto A, Tanimura S, Ohtsubo M, Misawa H, Miyazaki T, Leonor N, Taniguchi T, Fujita T, Kanekura Y, Komiya S, Yoshimura A: A new protein containing an SH2 domain that inhibits JAK kinases. *Nature* 1997; **387**: 921–924
7. Wakioka T, Sasaki A, Kato R, Shouda T, Matsumoto A, Miyoshi K, Tsuneoka M, Komiya S, Baron R, Yoshimura A: Spred, a Sprouty-related suppressor of Ras signaling. *Nature* 2001; **412**: 647–651
8. Kubo M, Hanada T, Yoshimura A: Suppressors of cytokine signaling and immunity. *Nature Immunol* 2003; **4**: 1169–1176
9. Naka T, Fujimoto M, Tsutsui H, Yoshimura A: Regulation of cytokine and TLR signalings by SOCS and others. *Adv Immunol* 2005; **87**: 61–122
10. Yoshimura A, Naka T, Kubo M: SOCS proteins, cytokine signaling and immune regulation. *Nat Rev Immunol* 2007; **7**: 454–465
11. Kamizono S, Hanada T, Yasukawa H, Minoguchi S, Kato R, Minoguchi M, Hattori K, Morita S, Kitamura T, Kato H, Nakayama K, Yoshimura A: The SOCS box of SOCS-1 accelerates ubiquitin-dependent proteolysis of TEL-JAK2. *J Biol Chem* 2001; **276**: 12530–12538
12. Yasukawa H, Misawa H, Sakamoto H, Masuhara M, Sasaki A, Wakioka T, Ohtsuka A, Imaizumi T, Matsuda T, Ihle JN, Yoshimura A

- ra A: The JAK-Binding Protein JAB Inhibits Janus Tyrosine Kinase Activity Through Binding in the Activation Loop. *EMBO J* 1999; **18**: 1309–1320
13. Sasaki A, Yasukawa H, Shouda T, Kitamura T, Dikic I, Yoshimura A: CIS3/SOCS3 suppresses erythropoietin signaling by binding the EPO receptor and JAK2. *J Biol Chem* 2000; **275**: 29338–29347
  14. Behrmann I, Tsiaris W, Sasaki A, Schneider-Mergener J, Yoshimura A, Neel BG, Heinrich PC, Schaper F: SHP2 and SOCS3 contribute to Y759-dependent attenuation of IL-6-signaling through gp130. *J Biol Chem* 2003; **278**: 661–671
  15. Yasukawa H, Ohishi M, Mori H, Murakami M, Chinen T, Aki D, Hanada T, Takeda K, Akira S, Hoshijima M, Hirano T, Chien KR, Yoshimura A: IL-6 induces an anti-inflammatory response in the absence of SOCS3 in macrophages. *Nature Immunol* 2003; **4**: 551–556
  16. Sasaki A, Yasukawa H, Suzuki A, Kamizono S, Syoda T, Kinjyo I, Sasaki M, Johnston JA, Yoshimura A: Cytokine-inducible SH2 protein-3 (CIS3/SOCS3) inhibits Janus tyrosine kinase by binding through the N-terminal kinase inhibitory region as well as SH2 domain. *Genes Cells* 1999; **4**: 339–351
  17. Marine J-C, Topham DJ, McKay C, Wang D, Parganas E, Stravopodis D, Yoshimura A, Ihle JN: SOCS1 Deficiency Causes a Lymphocyte-Dependent Perinatal Lethality. *Cell* 1999; **98**: 609–616
  18. Torisu T, Nakaya M, Watanabe S, Hashimoto M, Yoshida H, Chinen T, Yoshida R, Okamoto F, Hanada T, Torisu K, Takaesu G, Kobayashi T, Yasukawa H, Yoshimura A: Suppressor of cytokine signaling 1 protects mice against concanavalin A-induced hepatitis by inhibiting apoptosis. *Hepatology* 2008; **47**: 1644–1654
  19. Horino J, Fujimoto M, Terabe F, Serada S, Takahashi T, Soma Y, Tanaka K, Chinen T, Yoshimura A, Nomura S, Kawase I, Hayashi N, Kishimoto T, Naka T: Suppressor of cytokine signaling-1 ameliorates dextran sulfate sodium-induced colitis in mice. *Int Immunol* 2008; **20**: 753–762
  20. Tanaka K, Ichiyama K, Hashimoto M, Yoshida H, Takimoto T, Takaesu G, Torisu T, Hanada T, Yasukawa H, Fukuyama S, Inoue H, Nakanishi Y, Kobayashi T, Yoshimura A: Loss of Suppressor of Cytokine Signaling 1 in Helper T Cells Leads to Defective Th17 Differentiation by Enhancing Antagonistic Effects of IFN-gamma on STAT3 and Smads. *J Immunol* 2008; **180**: 3746–3756
  21. Takahashi Y, Dominici M, Swift J, Nagy C, Ihle JN: Trophoblast stem cells rescue placental defect in SOCS3-deficient mice. *J Biol Chem* 2006; **281**: 11444–11445
  22. Kimura A, Kinjyo I, Matsumura Y, Mori H, Mashima R, Harada M, Chien KR, Yasukawa H, Yoshimura A: SOCS3 is a physiological negative regulator for granulopoiesis and G-CSF receptor signaling. *J Biol Chem* 2004; **279**: 6905–6910
  23. Lee CK, Raz R, Gimeno R, Gertner R, Wistinghausen B, Takeshita K, DePinho RA, Levy DE: STAT3 is a negative regulator of granulopoiesis but is not required for G-CSF-dependent differentiation. *Immunity* 2002; **17**: 63–72
  24. Mori H, Hanada R, Hanada T, Aki D, Mashima R, Nishinakamura H, Torisu T, Chien KR, Yasukawa H, Yoshimura A: SOCS3 deficiency in the brain elevates leptin sensitivity and confers resistance to diet-induced obesity *Nature Medicine* 2004; **10**: 739–743
  25. Kievit P, Howard JK, Badman MK, Balthasar N, Coppari R, Mori H, Lee CE, Elmquist JK, Yoshimura A, Flier JS: Enhanced leptin sensitivity and improved glucose homeostasis in mice lacking suppressor of cytokine signaling-3 in POMC-expressing cells. *Cell Metab* 2006; **4**: 123–132
  26. Shi H, Tzamelis I, Bjorbaek C, Flier JS: Suppressor of cytokine signaling 3 is a physiological regulator of adipocyte insulin signaling. *J Biol Chem* 2004; **279**: 34733–34740
  27. Torisu T, Sato N, Yoshiga D, Kobayashi T, Yoshioka T, Mori H, Iida M, Yoshimura A: The dual function of hepatic SOCS3 in insulin resistance *in vivo*. *Genes Cells* 2007; **12**: 143–154.
  28. Kinjyo I, Hanada T, Inagaki-Ohara K, Mori H, Aki D, Ohishi M, Yoshida H, Kubo M, Yoshimura A: SOCS1/JAB Is a Negative Regulator of LPS-Induced Macrophage Activation. *Immunity* 2002; **17**: 583–591
  29. Mansell A, Smith R, Doyle SL, Gray P, Fenner JE, Crack PJ, Nicholson SE, Hilton DJ, O'Neill LA, Hertzog PJ: Suppressor of cytokine signaling 1 negatively regulates Toll-like receptor signaling by mediating Mal degradation. *Nat Immunol* 2006; **7**: 148–155
  30. Kimura A, Naka T, Muta T, Takeuchi O, Akira S, Kawase I, Kishimoto T: Suppressor of cytokine signaling-1 selectively inhibits LPS-induced IL-6 production by regulating JAK-STAT. *Proc Natl Acad Sci USA* 2005; **102**: 17089–17094
  31. Hanada T, Yoshida H, Kato S, Tanaka K, Masutani K, Tsukada J, Nomura Y, Mimata H, Kubo M, Yoshimura A: Suppressor of cytokine signaling-1 is essential for suppressing dendritic cell activation and systemic autoimmunity. *Immunity* 2003; **19**: 437–450
  32. Evel-Kabler K, Song XT, Aldrich M, Huang XF, Chen SY: SOCS1 restricts dendritic cells' ability to break self tolerance and induce antitumor immunity by regulating IL-12 production and signaling. *J Clin Invest* 2006; **116**: 90–100
  33. Hanada T, Tanaka K, Matsumura Y, Yamauchi M, Nishinakamura H, Aburatani H, Mashima R, Kubo M, Kobayashi T, Yoshimura A: Induction of hyper Th1 cell-type immune responses by dendritic cells lacking the suppressor of cytokine signaling-1 gene. *J Immunol* 2005; **174**: 4325–4332
  34. Shen L, Evel-Kabler K, Strube R, Chen SY: Silencing of SOCS1 enhances antigen presentation by dendritic cells and antigen-specific anti-tumor immunity. *Nat Biotechnol* 2004; **22**: 1546–1553
  35. Chinen T, Kobayashi T, Ogata H, Takaesu G, Takaki H, Hashimoto M, Yagita H, Nawata H, Yoshimura A: Suppressor of Cytokine Signaling-1 Regulates Inflammatory Bowel Disease in Which Both IFN-gamma and IL-4 Are Involved. *Gastroenterology* 2006; **130**: 373–388
  36. Matsumura Y, Kobayashi T, Ichiyama K, Yoshida R, Hashimoto M, Takimoto T, Tanaka K, Chinen T, Shichita T, Wyss-Coray T, Sato K, Yoshimura A: Selective expansion of foxp3-positive regulatory T cells and immunosuppression by suppressors of cytokine signaling 3-deficient dendritic cells. *J Immunol* 2007; **179**: 2170–2179
  37. Kinjyo I, Inoue H, Hamano S, Fukuyama S, Yoshimura T, Koga K, Takaki H, Himeno K, Takaesu G, Kobayashi T, Yoshimura A: Loss of SOCS3 in T helper cells resulted in reduced immune responses and hyperproduction of interleukin 10 and transforming growth factor-beta 1. *J Exp Med* 2006; **203**: 1021–1031
  38. Ogata H, Chinen T, Yoshida T, Kinjyo I, Takaesu G, Shiraishi H, Iida M, Kobayashi T, Yoshimura A: Loss of SOCS3 in the liver promotes fibrosis by enhancing STAT3-mediated TGF-beta1 production. *Oncogene* 2006; **25**: 2520–2530
  39. Li Y, Chu N, Rostami A, Zhang GX: Dendritic cells transduced with SOCS-3 exhibit a tolerogenic/DC2 phenotype that directs type 2 Th cell differentiation *in vitro* and *in vivo*. *J Immunol* 2006; **177**: 1679–1688
  40. Suzuki A, Hanada T, Mitsuyama K, Yoshida T, Kamizono S, Hoshino T, Kubo M, Yamashita A, Okabe M, Takeda K, Akira S, Matsumoto S, Toyonaga A, Sata M, Yoshimura A: CIS3/SOCS3/SSI3 Plays a Negative Regulatory Role in STAT3 Activation and Intestinal Inflammation. *J Exp Med* 2001; **193**: 471–482
  41. Shouda T, Yoshida T, Hanada T, Wakioka T, Oishi M, Miyoshi K, Komiya S, Kosai K, Hanakawa Y, Hashimoto K, Nagata K, Yoshimura A: Induction of the cytokine signal regulator SOCS3/CIS3 as a therapeutic strategy for treating inflammatory arthritis. *J Clin Invest* 2001; **108**: 1781–1788
  42. Chen Z, Laurence A, Kanno Y, Pacher-Zavisin M, Zhu BM, Tato C, Yoshimura A, Hennighausen L, O'Shea JJ: Selective regulatory function of Soes3 in the formation of IL-17-secreting T cells. *Proc Natl Acad Sci USA* 2006; **103**: 8137–8142

43. Seki Y, Inoue H, Nagata N, Hayashi K, Fukuyama S, Matsumoto K, Komine O, Hamano S, Himeno K, Inagaki-Ohara K, Cacalano N, O'Garra A, Oshida T, Saito H, Johnston JA, Yoshimura A, Kubo M: SOCS-3 regulates onset and maintenance of TH2-mediated allergic responses. *Nature Medicine* 2003; **9**: 1047–1054
44. Yoshida T, Ogata H, Kamio M, Joo A, Shiraishi H, Tokunaga Y, Sata M, Nagai H, Yoshimura A: SOCS1 is a suppressor of liver fibrosis and hepatitis-induced carcinogenesis. *J Exp Med* 2004; **199**: 1701–1707
45. Ogata H, Kobayashi T, Chinen T, Takaki H, Sanada T, Minoda Y, Koga K, Takaesu G, Maehara Y, Iida M, Yoshimura A: Deletion of the SOCS3 gene in liver parenchymal cells promotes hepatitis-induced hepatocarcinogenesis. *Gastroenterology* 2006; **131**: 179–193
46. Hanada T, Kobayashi T, Chinen T, Saeki K, Takaki H, Koga K, Minoda Y, Sanada T, Yoshioka T, Mimata H, Kato S, Yoshimura A: IFN $\gamma$ -dependent, spontaneous development of colorectal carcinomas in SOCS1-deficient mice. *J Exp Med* 2006; **203**: 1391–1397
47. Yoshimura A: Signal transduction of inflammatory cytokines and tumour development. *Cancer Sci* 2006; **97**: 439–447
48. Sasaki A, Taketomi T, Wakioka T, Kato R, Yoshimura A: Identification of a dominant negative form of Sproutys that potentiates FGF- but not EGF-induced ERK activation. *J Biol Chem* 2001; **276**: 36804–36808
49. Kato R, Nonami A, Taketomi T, Wakioka T, Kuroiwa A, Matsuda Y, Yoshimura A: Molecular cloning of mammalian Spred-3 which suppresses tyrosine kinase-mediated Erk activation. *Biochem Biophys Res Commun* 2003; **302**: 767–772
50. Sasaki A, Taketomi T, Kato R, Saeki K, Nonami A, Sasaki M, Kuriyama M, Saito N, Shibuya M, Yoshimura A: Mammalian Sprouty4 suppresses Ras-independent ERK activation by binding to Raf1. *Nature Cell Biol* 2003; **5**: 427–432
51. Taketomi T, Yoshiga D, Taniguchi K, Kobayashi T, Nonami A, Kato R, Sasaki M, Sasaki A, Ishibashi H, Moriyama M, Nakamura KI, Nishimura J, Yoshimura A: Loss of mammalian Sprouty2 leads to enteric neuronal hyperplasia and esophageal achalasia. *Nature Neurosci* 2005; **8**: 855–857
52. Taniguchi K, Ayada T, Ichiyama K, Kohno R, Yonemitsu Y, Minami Y, Kikuchi A, Maehara Y, Yoshimura A: Sprouty2 and Sprouty4 are essential for embryonic morphogenesis and regulation of FGF signaling. *Biochem Biophys Res Commun* 2007; **352**: 896–902
53. Nonami A, Kato R, Taniguchi K, Yoshiga D, Taketomi T, Fukuyama S, Harada M, Sasaki A, Yoshimura A: Spred-1 negatively regulates interleukin-3-mediated ERK/mitogen-activated protein (MAP) kinase activation in hematopoietic cells. *J Biol Chem* 2004; **279**: 52543–52551
54. Inoue H, Kato R, Fukuyama S, Nonami A, Taniguchi K, Matsumoto K, Nakano T, Tsuda M, Matsumura M, Kubo M, Ishikawa F, Moon BG, Takatsu K, Nakanishi Y, Yoshimura A: Spred-1 negatively regulates allergen-induced airway eosinophilia and hyperresponsiveness. *J Exp Med* 2005; **201**: 73–82
55. Nobuhisa I, Kato R, Inoue H, Takizawa M, Okita K, Yoshimura A, Taga T: Spred-2 Suppresses Aorta-Gonad-Mesonephros Hematopoiesis by Inhibiting MAP Kinase Activation. *J Exp Med* 2004; **199**: 737–742
56. Nonami A, Taketomi T, Kimura A, Saeki K, Takaki H, Sanada T, Taniguchi K, Harada M, Kato R, Yoshimura A: The Sprouty-related protein, Spred-1, localizes in a lipid raft/caveola and inhibits ERK activation in collaboration with caveolin-1. *Genes Cells* 2005; **10**: 887–895
57. Taniguchi K, Kohno R, Ayada T, Kato R, Ichiyama K, Morisada T, Oike Y, Yonemitsu Y, Maehara Y, Yoshimura A: Spreds are essential for embryonic lymphangiogenesis by regulating vascular endothelial growth factor receptor 3 signaling. *Mol Cell Biol* 2007; **27**: 4541–4550. Epub 2007 Apr 16
58. Wang S, Aurora AB, Johnson BA, Qi X, McAnally J, Hill JA, Richardson JA, Bassel-Duby R, Olson EN: The endothelial-specific microRNA miR-126 governs vascular integrity and angiogenesis. *Dev Cell* 2008; **15**: 261–271
59. Bentires-Alj M, Kontaridis MI, Neel BG: Stops along the RAS pathway in human genetic disease. *Nat Med* 2006; **12**: 283–285
60. Brems H, Chmara M, Sahbatou M, Denayer E, Taniguchi K, Kato R, Somers R, Messiaen L, De Schepper S, Fryns JP, Cools J, Marynen P, Thomas G, Yoshimura A, Legius E: Germline loss-of-function mutations in SPRED1 cause a neurofibromatosis 1-like phenotype. *Nature Genet* 2007; **39**: 1120–1126. Epub 2007 Aug 19
61. Yoshida T, Hisamoto T, Akiba J, Koga H, Nakamura K, Tokunaga Y, Hanada S, Kumemura H, Maeyama M, Harada M, Ogata H, Yano H, Kojiro M, Ueno T, Yoshimura A, Sata M: Spreds, inhibitors of the Ras/ERK signal transduction, are dysregulated in human hepatocellular carcinoma and linked to the malignant phenotype of tumours. *Oncogene* 2006; **25**: 6056–6066
62. Miyoshi K, Wakioka T, Nishinakamura H, Kamio M, Yang L, Inoue M, Hasegawa M, Yonemitsu Y, Komiya S, Yoshimura A: The Sprouty-related protein, Spred, inhibits cell motility, metastasis, and Rho-mediated actin reorganization. *Oncogene* 2004; **23**: 5567–5576